Journal of Plant Physiology and Breeding

2014, 4(1): 13-21 ISSN: 2008-5168



# Effect of Salinity on Morpho-Physiological Characteristics of Spring Wheat Genotypes

### Seyed Ahmad Sadat Noori<sup>\*</sup>, Ali Izadi-Darbandi and Seyed Mohammad Mehdi Mortazavian

Received: April 16, 2013 Accepted: May 6, 2014

Department of Agronomy and Plant Breeding, College of Abouraihan, University of Tehran, Tehran Iran \*Corresponding author: e-mail: <u>noori@ut.ac.ir</u>

#### Abstract

Bread wheat germplasm tolerant to salinity with high end-use quality is required to maintain grain production in saline lands. Four spring bread wheat cultivars with different tolerance to salinity and their three  $F_3$  progenies were evaluated at four levels of salt concentrations; 0, 150, 200 and 250 Mm NaCl. Na<sup>+</sup>, K<sup>+</sup> and Cl<sup>-</sup> concentrations in the wheat penultimate leaf and some of the biomass and yield related traits were measured. The allelic variations of *Glu-1* loci in the crosses were examined. Salinity had a significant positive effect on Na<sup>+</sup> and Cl<sup>-</sup> concentrations and the Na<sup>+</sup>/K<sup>+</sup> ratio. As salinity level increased, yield and 1000 grain weight and K<sup>+</sup> concentration were declined. Most of the F<sub>3</sub> progenies did not show much improvement in terms of biomass or yield related traits. However, F<sub>3</sub> progenies of the Cajema×Lerma Rojo cross showed an improved quality score.

Keywords: Quality; Salinity; Wheat; Yield

### Introduction

With food productivity decreasing due to the deleterious effects of stressful agricultural conditions such as drought, cold and salinity (Mahajan and Tuteja 2005), minimizing yield losses is a major area of concern for all nations to cope with the increasing food requirements. Bread wheat (Triticum aestivum L.), along with other important cereal crops, constitutes a major portion of the total calorie requirements to most people worldwide, particularly to those residing in developing countries. Plant-base combat (i.e., developing saline resistant cultivars) with saline conditions seems to be the most promising way in minimizing the yield losses due to salinity (Harati and Noori 2005). However, breeding for adaptation to abiotic stress conditions is challenging because of the complexity of (i) the target environments and (ii) the adaptive mechanisms adopted by plants when exposed to the stressful conditions (Reynolds et al. 2005). Plant responses to salinity occur in two phases: a rapid osmotic phase that inhibits growth of young leaves and a slower ionic phase that accelerates senescence of mature leaves (Munns *et al.* 2002). The direct selection of superior salt-tolerant genotypes under field conditions is hindered by the significant influence that environmental factors have on the response of plants to salinity (Yamaguchi and Blumwald 2005).

In plants, increases of salt concentrations affect nearly all the major physiological processes including photosynthesis, protein synthesis, energy metabolism and lipid metabolism (Parida and Das 2005). The sensitivity of rice crop to salinity was attributed to the plant's inability to keep sodium (Na<sup>+</sup>) and chloride (Cl<sup>-</sup>) ions out of the transpiration stream (Hollington 1998). While increases in leaf Na<sup>+</sup> concentrations may help to maintain plant turgor, Na<sup>+</sup> cannot completely substitute for K<sup>+</sup> which is specifically required for protein synthesis and enzyme activation (Marschner 1988). Maintenance of adequate levels of K<sup>+</sup> is essential for plant survival in saline habitats. High K<sup>+</sup> concentrations in the stroma are necessary for the maintenance of optimum photosynthetic capacity under stress conditions (Chow et al. 1990). Under saline-sodic or sodic conditions, high levels of external Na<sup>+</sup> not only interfere with K<sup>+</sup> acquisition by the roots, but also may disrupt the integrity of root membranes and alter their selectivity. Application of external K<sup>+</sup> fertilizer resulted in increases in corn yield grown in sandy soil irrigated with increasing levels of saline water, though not proportional to the salty irrigating waters (Bar-Tal et al. 1991). The authors, then, concluded that other than its beneficial effects on increasing Na<sup>+</sup>/K<sup>+</sup> ratio in the plant cells, fertilization with K<sup>+</sup> did not reduce the deleterious effects of salinity. Chilling and salinity stresses have severely reduced 1000 grain weight in durum wheat (Katerji et al. 2005). They successfully improved 1000-grain weight, ash content and ßcarotene content of their durum wheat germplasm. However, the main parameters explaining gluten indices declined considerably in their durum wheat progenies.

The major class of glutenin polypeptides have been identified in wheat endosperm, designated as HMW-GS and LMW-GS; both classes occur in flour as cross-linked proteins, resulting from inter-polypeptide disulphide linkage. These genes coding for HMW-GS (*Glu-*1) and LMW-GS (*Glu-3*) are located respectively on the long and short arms of 1A, 1B and 1D chromosomes (Payne *et al.* 1980; Gupta *et al.* 1991). It is generally accepted that there are additive effect and epistasis interaction between glutenin subunits for bread making quality (Gupta *et al.* 1989; Nieto-Taladriz *et al.* 1994). The relationship between protein composition and bread making quality showed that the quantities of total flour protein, albumin+ globulin, and high and low molecular weight glutenin subunits in flour were significantly and positively correlated with bread loaf volume. However, the ratio of high to low molecular weight glutenin subunits had little association with loaf volume (Wang *et al.* 2007).

In this work, the morpho-physiological characteristics of the progenies of three wheat crosses along with their parental lines were examined. We studied the effect of three levels of salinity along with an untreated control and reported the ionic concentrations (Na<sup>+</sup>, K<sup>+</sup> and Cl) in the prependecular leaf of the bread wheat. Of particular interest, we also examined the transgressive segregation of alleles responsible for quality within these crosses wheat and interestingly, observed transgressive segregant progenies of one of the crosses with improved quality score (Payne et al. 1981).

### **Materials and Methods**

# Germplasm development and experimental design

In this study, we examined the ionic concentrations effects on yield components of the parental lines and the progenies of the test crosses obtained from a common moderate salt-tolerant spring wheat cultivar from CIMMYT- Mexico "Cajema" as the common parent and three other parents including "Sette Cerros", Lermaroja (both moderate salt tolerant cultivars from CIMMYT-Mexico), and Ho<sub>2</sub> (a moderate salt tolerant

cultivar from Libya). We also screened the parental lines and the progenies to investigate the transgressive segregations in allelic variations in the germplasm studied. The crosses were conducted at Abouraihan Campus, University of Tehran. For morphological and quality scoring determinations, F<sub>3</sub> generation of these test crosses and the parental lines were used along with one check cultivar (a British salt-sensitive) - "Axona". assessing morphological For and mineral concentrations, a factorial experiment with two factors, i.e. germplasm and salinity, was employed. The germplasm factor included (i) the four parental lines, (ii) the three test crosses at F<sub>3</sub> generation, and (iii) the salt-sensitive check cultivar (eight in total). The salinity factor consisted of four levels of salty water, i.e. 0 mM, 150 mM, 200 mM and 250 mM. We conducted the experiment in a randomized complete block design with three replications. After normalizing the data by square root transformation, the GLM procedure was used for statistical tests of significance. Dancan Multiple Range Test (DMRT) was used for mean comparisons.

Growth condition and salinity stress imposition The experiment was conducted in a glasshouse with a day/night temperature of  $22\pm2$  °C/16 $\pm2$ °C. The plants were grown in natural daylight supplemented with 400 watt mercury vapor lamps for 16 h per day. We used river sand as potting medium. The sand was washed with tap water for one week and oven dried. Plastic pots of 18 cm diameter / 19 cm deep were filled with 4.40 kg oven-dried sand. The potting medium prior to the experiment was equilibrated with full strength nutrient solution (Hewitt *et al.* 1966). For salt stress imposition, we designated four irrigation regimes of full strength nutrient solution (Hewitt *et al.* 1966) each accompanied by NaCl at final concentrations of 0 mM (for control), 150, 200 and 250 mM. Seeds were germinated during six days and thereafter, five seedlings of each breeding material were transplanted in each pot  $(10\times10$ cm). Salt imposition was started 18 days after the experiment commencement and the concentrations were increased gradually by increasing increments of 25 mM every other day until the final treatment concentration was met. Additionally, twice per week, 400 ml of deionized water was added to each pot to maintain sand moisture and to prevent salt accumulation.

# Plant morphological and physiochemical characteristics

For plant characters we measured plant height (PH), spike length (SL), spike weight (SW), straw weight (STW), number of grains per spike (NGPE), grain yield per plant (GYPP) and 1000 kernel weight (1000 KW). For assessing sodium, potassium and chloride ionic contents in the leaves, the perpendicular leaf from 2-3 plants of each replication were removed 30 days after the beginning of the salt treatment (Chow *et al.* 1990; Harati and Sadat Noori 2005)

# Protein extraction and assessing banding patterns

For protein extraction, we employed the sequential extraction procedure described by Singh *et al.* (1991), with some modifications (Izadi-Darbandi *et al.* 2010). Identification of banding patterns of HMW-GS and giving quality scores were conducted according to Payne and Lawrence (cited in Payne *et al.* 1981).

### Salt effects on yield related traits

Salinity treatments decreased biomass and yield related traits in wheat germplasm, invariably. significant differences There was among genotypes for plant height, spike length and number of grains per spike. We observed considerable decreases in the biomass and yield associated traits in wheat parental lines and F<sub>3</sub> progenies. Analysis of variance (Table 1) revealed significant (p-value<0.0001) statistically difference amongst the four levels of salinity for plant height, straw weight, spike length, spike weight, number of grains per spike, grain yield per plant and 1000 grain weight. Decreases of biomass and yield components are hallmarks of growing crops in saline conditions and were repeatedly in the literature reported (Allakhverdiev et al. 2000; Reddy et al. 2003; Mahajan and Tuteja 2005; Farooq and Azam 2006; Soltani et al. 2006; Kara and Altindal 2011). Despite the prominent and significant differences observed in the levels of salinity for biomass and yield related traits, the germplasm revealed to have significant effect only on plant length (Table height and spike 1) (pvalue<0.0001) and on the number of grains per spike (p-value<0.05). Comparison of means indicated that NaCl concentration of 150 mM imposed considerable adverse effect on the

Sources of Variation	Salinity	Genotype	Salinity × Genotype	Error	R-Square (%)
df	3	7	21	64	
Biomass associated traits					
Plant height	6667.82***	215.94***	116.32**	36.29	91
Straw weight	5.13***	0.10	0.04	0.10	73
Yield related traits					
Spike length	43.14***	5.87***	1.17**	32.04	85
Spike weight	23.16***	0.06	0.08	14.42	83
Number of grains per spike	4316.24***	51.08*	24.59	21.34	91
Grain yield per plant	15.88***	0.05	0.03	0.08	91
1000 grain weight	5255.18***	34.55	31.84	2190.01	88
Physiochemical traits					
Chloride [Cl -]	23.017***	1.38	0.87	0.79	66
Sodium [Na+]	13.62***	1.47**	0.44	0.33	74
Potassium [K+]	4.26***	0.06	0.02	0.04	85
Na+ / K+ ratio	120.47***	7.68*	3.26	2.99	72

Table 1. Analysis of variance of some morpho-physiological traits of wheat genotypes under salinity stress.

\*: P< 0.05, \*\*: P< 0.01, \*\*\*: P< 0.001



Figure 1. The mean reduction in plant height, number of grains per plant, grain yield per plant and 1000 grain weight in wheat genotypes.

germplasm growth and for concentrations beyond 150 mM there was no considerable decrease. Figure 1 shows decreases observed in plant height, number of grains per plant, grain yield per plant and 1000 grain weight upon imposition of 150 mM NaCl on wheat germplasm. Accordance with other research reports (Francois et al. 1986; Katerji et al. 2005; Farooq and Azam 2006), salinity has shown a highly significant (pvalue<0.01) effect on yield components and increases in salt concentration reduced grain yield per plant and 1000 grain weight significantly.

# Salt effects on ion concentration

Analysis of variance showed significant effect of salinity on concentrations of chloride, sodium and and Na<sup>+</sup>/K<sup>+</sup> potassium ratio whereas, the germplasm showed only significant effect on sodium concentrations and  $Na^+/K^+$  ratio (Table 1). Upon imposition of salt stress by increasing concentration of NaCl from 0 mM to 150, 200 and 250 mM, we observed the sharpest changes of ionic concentrations occurred from 0 to 150 mM beyond which little or no significant change was detected in the ionic concentrations of chloride, sodium and potassium. In the tested germplasm, of NaCl increasing concentrations caused increases in Na<sup>+</sup> and Cl<sup>-</sup> accumulation and decreases in  $K^+$ concentrations in the prependecular leaf of wheat (Figure 2). To present the ionic concentrations in the normally grown wheat and salt stressed germplasm, we averaged the concentrations in all NaCl treated groups and showed them in a two groups of normal versus salt stressed groups (Figure 3). The results were in concordence with the report by Khatun and Flowers (1995) who showed the increase in Na<sup>+</sup>

Number of grains per spike

and Cl<sup>-</sup> concentrations and decrease in K<sup>+</sup> concentration upon NaCl stress. Negative correlation between Na<sup>+</sup> and K<sup>+</sup> was also postulated before (Marschner 1988; Harati and Noori 2005: Farooq and Azam 2006). Maintenance of low Na/K ratios is thought as one favorable index for salt tolerance (Zhu et al. 1998). Analysis of variance showed availability of genetic variation for this character among the germplasm tested. Cajema as a common parent showed the lowest amount of Na/K ratio and F1 generation between Cajema\*Ho2 got the highest related ratio. It seems that the salt tolrenace of Cajema was higher than its cross with Ho2.

### Allelic variations in *Glu-1* loci

Apart from the trends observed in yield related traits and changes reported in ionic concentrations in agreement with other previously reported studies, we also noticed a critical observation when profiled the allelic variations of *Glu-1* loci

in the parental lines and their progenies. We observed a transgressive segregation, occurred likely due to a crossing over, for improved quality score among  $F_3$  progenies of one of the wheat (Cajema×Lerma crosses Roja). Table 2 summarizes the allelic variations within the parental lines as well as the progenies in Glu-1 loci. Among evaluated germplasms, the Payne quality scores were 6 (low), 8 (moderate) and 10 (good). Glutenin profile of the parental lines Cajema and Lerma Roja and their hybrid (Cajema×Lerma Roja) is shown in Figure 3. The Glu-D1 allele in the progenies of Cajema×Lerma Roja was "5+10" alleles instead of their parental "2+12" and 17 instead of 7+8 and 17+18 alleles in Glu-B1. This observation can be explained by a crossing over occurred in this cross that adds up the quality score.

Germplasm	Glu-A1	Glu-B1	Glu-D1	Payne Scores
Cajema×Sette Cerros	2*	7+8	2+12	8
Cajema×Ho2	1	7+8	2+12	8
Cajema×Lermaroja	2*	17	5+10	10
Axona	Null	7+8	2+12	6
Sette Cerros	Null	7+8	2+12	6
Ho2	2*	17+18	2+12	8
Lermaroja	2*	17+18	2+12	8
Cajema	2*	7+8	2+12	8

Table 2. Allelic variation in *Glu-1* loci of wheat genotypes profiled on a SDS-PAGE pattern.



Figure 2. Changes in ionic concentration of Cl<sup>-</sup>, Na<sup>+</sup>, K<sup>+</sup> and Na<sup>+</sup> / K<sup>+</sup> ratio averaged over all wheat genotypes.



Figure 3. The means of Cl<sup>-</sup>, Na<sup>+</sup> and K+ concentrations as well as Na<sup>+</sup> / K<sup>+</sup> ratio in wheat genotypes. The concentrations are given in mgr  $100^{-1}$ gr dry matter.

Salinity had significant effect on concentrations of chloride, sodium and potassium and Na<sup>+</sup> / K<sup>+</sup> ratio. The mean values of  $F_3$  generations in terms of yield and biomass was not reduced compared to their respective parents. It is expected that we will, hopefully, find more potential of salt tolerant lines in the next

generations by transgressive segregation. It could be due to separation of the effective genes of salinity tolerance in  $F_3$  progenies. We found transgressive segregation in case of quality scores in  $F_3$  progenies in one of the wheat crosses (Cajema×Lerma Roja).



Figure 4. SDS-PAGE banding patterns of *Glu-1D* loci in the wheat genotypes (from left to right; A: Cajema×Ho2, B: Ho2, C: Cajema×Lerma Rojo, D: Lerma Rojo, E: No sample (control), F: Chines Spring, G: Cajema×Sette Cerros, H: Cajema, I: Sette Cerros and J: Axona).

# Acknowledgement

We would like to thank Prof. Bahman Ehdaie from Department of Botany and Plant Science, University of California for his numerous suggestions and helpful discussion and Dr. Mohsen Mohammadi from Seed and Plant Improvement Institute for his comments. Financial support for this research was granted by the University of Tehran and the Ministry of Science, Research and Technology of Iran which are gratefully acknowledged.

### References

- Allakhverdiev SI, Sakamoto A, Nishiyama Y, Inaba M and Murata N, 2000. Ionic and osmotic effects of NaCl-induced inactivation of photosystems I and II in Synechococcus sp. Plant physiology 123: 1047-1056.
- Bar-Tal A, Feigenbaum S and Sparks D, 1991. Potassium-salinity interactions in irrigated corn. Irrigation Science 12: 27-35.
- Chow W, Ball M and Anderson J, 1990. Growth and photosynthetic responses of spinach to salinity: implications of K+ nutrition for salt tolerance. Functional Plant Biology 17: 563-578.
- Farooq S and Azam F, 2006. The use of cell membrane stability (CMS) technique to screen for salt tolerant wheat varieties. Journal of Plant Physiology 163: 629-637.
- Francois L, Maas E, Donovan T and Youngs V, 1986. Effect of salinity on grain yield and quality, vegetative growth and germination of semi-dwarf and durum wheat. Agronomy Journal 78: 1053-1058.

Gupta R, Bekes F and Wrigley C, 1991. Prediction of physical dough properties from glutenin subunit

composition in bread wheats: correlation studies. Cereal Chemistry 68: 328-333.

- Gupta R, Singh N and Shepherd K, 1989. The cumulative effect of allelic variation in LMW and HMW glutenin subunits on dough properties in the progeny of two bread wheats. Theoretical and Applied Genetics 77: 57-64.
- Harati M and Sadat Noori SA, 2005. Breeding for salt-resistance using iransgressive segregation in spring wheat. Journal of Sciences, Islamic Republic of Iran 16: 217-222.
- Hewitt EJ, Bureaux CA and Royal F, 1966. Sand and Water Culture Methods Used in the Study of Plant Nutrition, Cambridge Univ Press.
- Hollington P, 1998. Technological breakthroughs in screening/breeding wheat varieties for salt tolerance. National Conference on "Salinity Management in Agriculture" December 2 - 5, CSSRI Karnal, India.
- Izadi-Darbandi A, Yazdi-Samadi B, Shanejat-Boushehri AA and Mohammadi M, 2010. Allelic variations in *Glu-1* and *Glu-3* loci of historical and modern Iranian bread wheat (*Triticum aestivum* L.) cultivars. Journal of Genetics 89: 193-199.
- Kara IaB and Altindal D, 2011. Effect of salinity (NaCl) on germination, seedling growth and nutrient uptake of different Triticale genotypes. Turkish Journal of Field Crops 16: 225-232.
- Katerji N, Van Hoorn J, Fares C, Hamdy A, Mastrorilli M and Oweis T, 2005. Salinity effect on grain quality of two durum wheat varieties differing in salt tolerance. Agricultural Water Management 75: 85-91.
- Khatun S and Flowers T, 1995. Effects of salinity on seed set in rice. Plant, Cell & Environment 18: 61-67.
- Mahajan S and Tuteja N, 2005. Cold, salinity and drought stresses: an overview. Archives of Biochemistry and Biophysics 444: 139-158.
- Marschner H, 1988. Mineral nutrition in higher plants. Plant, Cell and Environment 11: 147-148.
- Munns R, Husain S, Rivelli AR, James RA, Condon A, Lindsay MP, Lagudah ES, Schachtman DP and Hare RA, 2002. Avenues for increasing salt tolerance of crops, and the role of physiologically based selection traits. Plant and Soil 247: 93-105.
- Nieto-Taladriz M, Perretant M and Rousset M, 1994. Effect of gliadins and HMW and LMW subunits of glutenin on dough properties in the F 6 recombinant inbred lines from a bread wheat cross. Theoretical and Applied Genetics 88: 81-88.
- Parida AK and Das AB, 2005. Salt tolerance and salinity effects on plants: a review. Ecotoxicology and Environmental Safety 60: 324-349.
- Payne PI, Corfield KG, Holt LM and Blackman JA, 1981. Correlations between the inheritance of certain high-molecular weight subunits of glutenin and bread-making quality in progenies of six crosses of bread wheat. Journal of the Science of Food and Agriculture 32: 51-60.
- Payne P, Law C and Mudd E, 1980. Control by homoeologous group 1 chromosomes of the high-molecularweight subunits of glutenin, a major protein of wheat endosperm. Theoretical and Applied Genetics 58: 113-120.
- Reddy B, Sanjana Reddy P, Bidinger F and Blümmel M, 2003. Crop management factors influencing yield and quality of crop residues. Field Crops Research 84: 57-77.
- Reynolds M, Mujeeb-Kazi A and Sawkins M, 2005. Prospects for utilising plant-adaptive mechanisms to improve wheat and other crops in drought-and salinity-prone environments. Annals of Applied Biology 146: 239-259.
- Soltani A, Gholipoor M and Zeinali E, 2006. Seed reserve utilization and seedling growth of wheat as affected by drought and salinity. Environmental and Experimental Botany 55: 195-200.
- Wang Y, Khan K, Hareland G and Nygard G, 2007. Distribution of protein composition in bread wheat flour mill streams and relationship to breadmaking quality. Cereal Chemistry 84(3): 271-275.
- Yamaguchi T and Blumwald E, 2005. Developing salt-tolerant crop plants: challenges and opportunities. Trends in Plant Science 10: 615-620.
- Zhu JK, Liu J and Xiong L, 1998. Genetic analysis of salt tolerance in Arabidopsis: evidence for a critical role of potassium nutrition. The Plant Cell, Online 10: 1181-1192.