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Effect of 17β-estradiol on seedling and callus growth of German chamomile (*Matricaria chamomilla* L.)

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Abstract

To study the effect of 17β -estradiol on seedling growth, antioxidant enzyme activity and also, on callus induction from leaf explants of German Chamomile (*Matricaria chamomilla* L.), an experiment was conducted as a completely randomized design with three replications using MS medium containing different concentrations of 17β -estradiol (0, 0.01, 0.1, 1 and 10 mg/l) alone or in combination with 3 mg/l Benzylaminopurine (BAP) + 0 or 1.5 mg/l 1-naphthalene acetic acid (NAA). The results showed that 17β -estradiol at 0.01 and 0.1 mg/l increased root and shoot length and weight, respectively and at high concentration (10 mg/l) increased peroxidase, polyphenol oxidase and catalase activity of German chamomile seedlings. Also, the callus induction was observed after one to two weeks in all media, but the growth varied depending on the presence or absence of plant growth regulators and different concentrations of 17β estradiol. Maximum callus weight was obtained in 0.01 mg/l of 17β -estradiol with about three-fold increase in comparison with the control (MS without 17β -estradiol). This indicates that the 17-beta-estradiol at lower concentrations (0.01 mg/l) can significantly improve callus growth in the presence of plant growth regulators such as NAA and BAP. The results of this study indicate that using steroidal hormone 17β -estradiol can be used to optimize German chamomile cell growth under *in vitro* conditions.

Keywords: Antioxidant enzymes, Callus induction, In vitro culture, Steroid hormone

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Introduction

17β-estradiol, a steroid hormone with low molecular weight has key roles in controlling growth, development, reproduction and control of protein and mineral metabolism in mammals. In recent years, this hormone has been recognized in many plant species and extracted from tissues and organs such as roots, leaves and flowers (Janeczko and Skoczowski 2005; Genisel *et al.* 2013). Recent studies on the effects of this hormone have shown that its exogenic use in plants, stimulate cell division, root and shoot growth, embryo development, pollen tube growth, flowering, proliferation and callus growth and increase the activity of antioxidant enzymes (Erdal 2012a; Erdal and Dumlupinar 2011b; Genisel *et al.* 2013). It has also been reported that 17 β -estradiol is effective in callus induction and proliferating in *Polygonatum verticillatum* L. (Janeczko and Szybka 2001). Furthermore, the application of β estradiol leads to a significant increase in plant growth, protein and soluble sugars contents and antioxidant enzymes activity such as superoxide dismutase, peroxidase and catalase in *Cicer* arietinum L. (Erdal and Dumlupinar 2011a) and wheat seedlings under salt stress (Erdal 2012b). It has also been reported that this hormone can improve sugar, proline, protein, chlorophyll, glutathione and dry matter contents in wheat seedlings under salt stress conditions as compared with the control plants. Furthermore, lipid peroxidation and production of hydrogen peroxide and superoxide content induced by salt stress are decreased in wheat by using this hormone (Erdal 2012b). The effects of the 17 β -estradiol hormone on plant life have not been fully studied. Recent research shows the existence and effects of this hormone on plant growth under *in vitro* conditions (Janeczko 2012).

It was shown that in vitro callus induction and shoot regeneration in plants are affected by the application of proper plant growth regulators (Fazeli Behgo and Alizadeh Ajirlo 2015). Due to the lack of adequate research on the effect of this hormone on medicinal herbs, this study aimed to investigate the effect of mammalian sex hormones on callus and seedling growth and antioxidant in German enzymes activity chamomile (Matricaria chamomilla L.) under in vitro culture. German chamomile from the Asteraceae family is widely used in the pharmacopeia of many countries in the world due to its essential oils and secondary metabolites with pharmaceutical importance (Petronilho et al. 2012). In a previous study (Koohi et al. 2014), we investigated the callus induction from leaf explants of German chamomile on MS medium supplemented with different concentrations of plant growth regulators, and it was concluded that 1.5 mg/l NAA and 3 mg/l BAP is the most proper medium

for the callus induction. So, the main aim of the present study was to investigate the effect of different concentrations of 17β -estradiol on callus growth, antioxidant enzymes activity and plantlet growth of German chamomile under the *in vitro* culture conditions in the presence or absence of these phytohormones.

Materials and Methods

The chamomile seeds provided from Pakanbazr Co. (http://www.pakanbazr.com/), were washed with distilled water, sterilized for 10 minutes in 2.5% sodium hypochlorite and then were immersed for 30 seconds in 70% ethanol. Then, the seeds were rinsed three times with sterilized distilled water and were cultured in vessels containing MS medium (Murashige and Skoog 1962) in a growth chamber at 25±2 °C and the photoperiod of 16/8 h light and darkness, respectively. After germination, 10 days old seedlings were transferred to an MS medium containing different concentrations (0, 0.01, 0.1, 1 and 10 mg/l) of 17β-estradiol (Sigma-Aldrich, E8875). Due to the water insolubility of 17β estradiol in ethanol, it was first dissolved in one drop of 96% ethanol then the required distilled water was added to reach the proper concentration. It should be noticed that for negligible adverse effects of ethanol, it was added to the control treatment.

After 40 days, shoot and root length and weight and the activity of antioxidant enzymes were measured. To measure the activity of antioxidant enzymes including catalase (CAT), polyphenol oxidase (PPO) and peroxidase (POX), leaf tissues from 40-day-old seedlings were ground in liquid nitrogen and homogenized in 50 mM Tris-HCl buffer. The enzymes were extracted according to Sudhakar *et al.* (2001). After centrifugation at 12000 rpm for 20 min at 4 °C, the supernatant was taken to measure the activity of peroxidase and polyphenol oxidase based on Kar and Mishra (1976) and catalase activity according to Chance and Maehly (1955), by recoding absorbance at 425, 420 and 240 nm, respectively.

In addition, the effect of the 17β -estradiol hormone on callus induction and growth from leaf explants of chamomile was studied on MS medium containing different concentrations (0, 0.01, 0.1, 1, 10 mg/l) of this hormone alone or in combination with 3 mg/l BAP + 0 or 1.5 mg/l NAA according to Koohi *et al.* (2014). Five weeks after culture, callus induction rate and callus fresh weight (per explant) was recorded. Analysis of variance and comparison of means was done with the SAS software (Version 9.1). The data were reported as the mean ± standard error (SE) based on three replications.

Results

Seedling growth

The *in vitro* growth of German chamomile seedlings and the calluses induced from its leaf explants under different concentration of 17βestradiol are shown in Figure 1. The effect of 17βestradiol on all of the seedling characteristics was significant ($p \le 0.05$). The 17β-estradiol hormone was effective in increasing the shoot and root length of German chamomile seedlings. The highest shoot length and weight of chamomile seedlings were observed at 0.1 mg/l of 17βestradiol. However, the highest concentration significantly differed from the control regarding these characteristics (Figure 2). The root length and weight of German chamomile seedlings were also significantly increased at 0.01 and 0.1 mg/l of 17β -estradiol as compared to the control, but at higher concentrations (1 and 10 mg/l) its difference with the control was not significant.

On the other hand, the activity of peroxidase, polyphenol oxidase and catalase enzymes increased as the concentration of 17β -estradiol hormone was increased, so the maximum activity was observed at 10 mg/l concentration of this hormone. The lowest concentration of 17β estradiol (0.01 mg/l) was not significantly different from the control concerning all antioxidant enzymes activities (Figure 3).

Callus induction

Callus induction from leaf explants of the German chamomile in all media containing various concentrations of 17B-estradiol alone, or in combination with NAA (0 and 1.5 mg/l) and BAP (3 mg/l) was observed after two weeks. However, callus growth varied depending on the concentration of the steroid hormone and the presence or absence of phytohormones (Figure 4). On the MS medium containing 3 mg/l BAP plus 0 or 1.5 mg/l NAA, all of the leaf explants induced callus. The highest callus induction on MS medium containing 17B-estradiol without the phytohormones was only 37%, however, it was significantly greater than the control. On the MS medium containing both NAA and BAP, the addition of 17β -estradiol increased the callus weight. The maximum callus weight was obtained

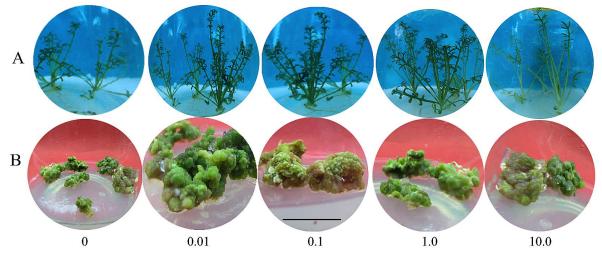


Figure 1. *In vitro* growth of German chamomile seedlings (A) and induced calluses from its leaf explants on MS medium containing different concentrations of 17β -estradiol (0, 0.01, 0.1, 1 and 10 mg/l) in combination with a 1.5 mg/l NAA and 3 mg/l BAP (B). Scale bar 50 mm.

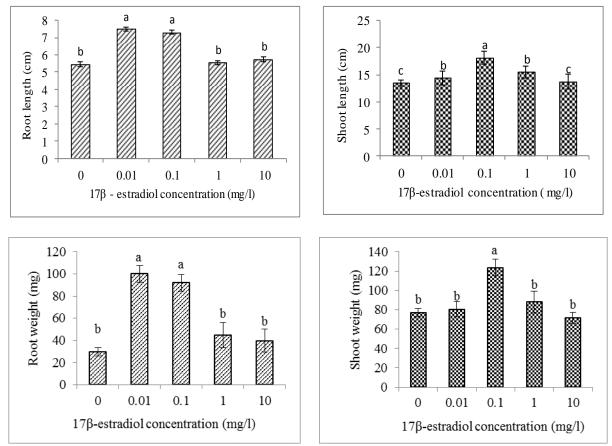


Figure 2. Effects of 17β -estradiol treatment on root and shoot length and weight of German chamomile seedlings. The error bars represent standard errors; Means with different letters are significantly different at $p \le 0.05$.

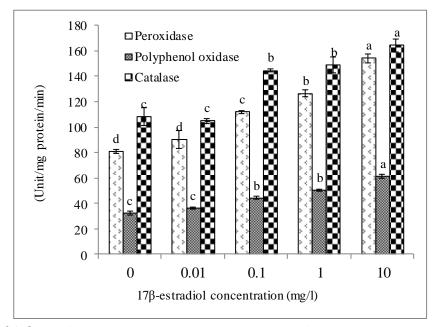


Figure 3. Effects of 17β -estradiol treatment on peroxidase, polyphenol oxidase and catalase enzymes activity of the German chamomile seedlings. The error bars represent standard errors; Means with different letters are significantly different ($p \le 0.05$) in each enzyme.

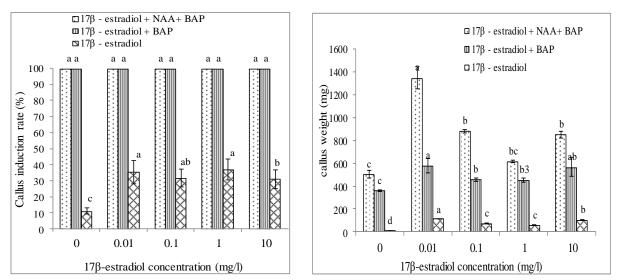


Figure 4. Callus fresh weight and induction rate (%) and German chamomile leaf explants in MS medium supplemented with NAA and BAP, MS medium with BAP only and MS medium without NAA and BAP under treatment with different concentrations of 17β -estradiol. In each trait, means with different letters show significant differences by the LSD test at the 5% probability level.

at 0.01 mg/l of 17β -estradiol with about three-fold increase. This indicates that the 17-beta-estradiol lower concentrations (0.01)at mg/l) can significantly improve callus growth in the presence of phytohormones such as NAA and BAP. Furthermore, on the MS medium supplemented with only BAP (3 mg/l without NAA) and various concentrations of 17βestradiol, the callus weight was significantly increased. So that, 17β -estradiol at 0.01 mg/l significantly increased callus weight in the auxinfree medium (about 50%) as compared with the medium consisted of BAP only. This represents an effective role of 17β-estradiol in improving the callus growth at the auxin-free medium. On the other hand, although leaf explants on MS medium without NAA or BAP, can produce callus as maximum as 37% as compared with the control (MS medium free of any hormones), the callus growth was much more reduced in comparison with the MS medium containing NAA and BAP. Maximum callus weight (114.6 mg) was obtained

on the MS medium containing 0.01 mg/l, which was significantly greater than other concentrations of 17β -estradiol (Figure 4).

Discussion

The results of this study about the effect of 17β estradiol on seedling growth of the German chamomile showed that at the low concentration of 17β -estradiol (0.01 and 0.1 mg/l), the root length and weight increased, and at 0.1 mg/l, the shoot length and weight were increased. It has been shown that external use of this steroid hormone significantly increases plant growth,

protein, soluble sugars and antioxidant enzymes (Erdal and Dumlupinar 2011a). Treatment of corn seedlings with this hormone under salinity stress has significantly alleviated the adverse effects of salinity (Erdal 2012c). In addition, the application of 17β -estradiol on wheat seedlings under salinity stress improved dry weight, glucose, proline, protein and chlorophyll contents (Erdal and Dumlupinar 2011b). Furthermore, spraying alfalfa (Medicago sativa L.) with nutrient solution containing 0.005 and 0.05 mg/l of 17B-estradiol and estrone increased root and shoot weight, but at 50 and 500 mg/l, the growth of this plant was inhibited (Shore et al. 1992). Some studies show that foliar application of hormones such as gibberellic acid, epibrassinolide and acetylsalicylic acid on plants under stress conditions can improve plant physiological and agro-morphological characteristics (Fathi et al. 2019; Pourasadollahi et al. 2019).

This 17β -estradiol hormone had a significant effect on the activity of antioxidant enzymes. The activity of peroxidase, polyphenol oxidase and catalase enzymes increased in the German chamomile seedlings. The highest activity of these enzymes was obtained at the highest concentration of 17β -estradiol (10 mg/l). Researches by Erdal (2012a, b) and Erdal and Dumlupinar (2011a) have also shown that the use of 17β -estradiol increases the activity of antioxidant enzymes. These enzymes provide tolerance and adaptation to environmental conditions mainly by purifying reactive oxygen species and reducing the harmful effects of stress on plants. Erdal and Dumlupinar (2011b) reported that this hormone increased concentrations of potassium, calcium, magnesium, manganese, aluminum, zinc, iron, copper, phosphorus and sulfur. It can be stated that this steroidal hormone, along with the increase of organic matter and minerals and the increase of antioxidant enzyme activity, improves the conditions for plant growth (Dumlupinar *et al.* 2011). The application of other steroidal hormones such as androsterone has also significantly increased the growth and activity of antioxidant enzymes in *Cicer arietinum* L. (Erdal and Dumlupinar 2011a).

As was shown, the highest callus weight of the German chamomile was obtained at the low concentration (0.01 mg/l) of 17β -estradiol. This suggests that 17β-estradiol can significantly increase the weight of callus in combination with NAA and BAP more than the amount reported previously in this plant in the presence of NAA and BAP (Koohi et al. 2014). In addition, increasing the callus weight in the auxin-free medium indicated the effective role of this hormone in improving callus growth conditions in the absence of auxin. Induction and growth of callus from leaf explants in the NAA-free medium seem to have been influenced by this steroidal hormone. In addition, callus induction of leaf explants the MS medium without in phytohormones showed that in the absence of BAP and NAA, it can induce callus formation in the leaf explants although, its growth was weak. Still, this hormone significantly increased callus growth if callus induction was made in the presence of BAP and NAA phytohormones. The results of the present study confirm the effective role of 17\beta-estradiol in the growth of calli

obtained from German chamomile, which was in agreement with the results of studies by Janeczko et al. (2002) that showed increased callus growth under the influence of 17B-estradiol and androstenedione. It has been shown that androstenedione at a rate of 1 µM induces callus growth from the scutellum of the immature wheat (Triticum aestivum L.) embryos. It has been also reported that 17β -estradiol at the concentration of 2-12 mg/l, increases the growth of Daucus carrota L. in the tissue culture medium by 100% (Janeczko and Skoczowski 2005). Janeczko and Szybka (2001) also showed that this hormone induces callus in Polygonatum verticillatum L.

Although the presence of steroid hormones in plants has been established (Geuns 1978; Simons and Grinwich 1989; Janeczko and Skoczowski 2005; Iino et al. 2007; Simersky et al. 2009; Janeczko 2012), little is known about the major role and effects of these hormones in plants. Similar to mammalian sex hormones, plant steroidal hormones play a key role in the flexibility of plants under varying environmental conditions. At the low concentrations, they affect plants by controlling cell division (lino et al. 2007). It was reported that when brassinosteroids bind to a receptor on the cell wall, they trigger a multi-level cascade of reactions that regulates the activity of several transcription factors especially CESTA (CES) (Khan et al. 2014). In animals, steroidal hormones are involved in regulating the transcription of specific genes through the plasma membrane and attaching to cytoplasmic or nuclear receptors. Milanesi et al. (2001) used the calli of Solanum glaucophyllum to study the expression of estrogen-binding proteins. They first used the

radioligand binding assay to investigate the specific binding site of 17β -estradiol and then used western blotting and ligand blotting methods to confirm the possible association of these loci with known isoforms. The calli of *Solanum glaucophyllum* not only contain 17β -estradiol and estrone hormones but also have abundant binding sites. These findings indicate the existence of estrogen receptor molecules in the plant. In addition, the presence of 17β -estradiol internal receptors was demonstrated in the pistil of *Gladiolus primulinus* (Janik and Adler 1984).

Production of plant secondary metabolites in cell and tissue culture of medicinal plants is a major goal in the production of medicinally important compounds using biotechnological tools. The first step to do this is the establishment of well-growing cells and calli under in vitro conditions. The optimization of cell growth in plant tissue culture results in better growth of cells and improved production of metabolites in bioreactors for industrial-scale production of active compounds. The results of this study indicate that steroidal hormone 17β-estradiol can be used to optimize the German chamomile cell growth under in vitro conditions. However, more study and further research are needed in this and other plants to reach to a decisive conclusion.

The low concentration of 17β -estradiol increased 17-est increased root and shoot weight of German chamomile seedlings and at higher concentrations increased antioxidant enzymes activities. Also, the highest callus weight of leaf explants was obtained with NAA and BAP phytohormones by adding low concentration of 17B-estradiol (0.01 mg/l). In the absence of phytohormones such as BAP and NAA, this hormone can induce callus formation in the leaf explants of this plant, but the growth rate of callus was much lower than that used in combination with BAP and NAA in the culture medium. Our study indicates that using steroidal hormone 17B-estradiol will be beneficial to optimize the German chamomile cells' growth under in vitro conditions.

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Conflict of Interest

The authors declare that they have no conflict of interest with any people or organization concerning the subject of the manuscript.

References

Chance B and Maehly AC, 1955. Assay of catalases and peroxidases. Methods in Enzymology 11: 764-755.

Dumlupinar R, Genisel M, Erdal S, Korkut T, Taspinar MS and Taskin M, 2011. Effects of progesterone, βestradiol and androsterone on the changes of inorganic element content in barley leaves. Biological Trace Element Research 143: 1740-1745.

Erdal S, 2012a. Exogenous mammalian sex hormones mitigate inhibition in growth by enhancing antioxidant activity and synthesis reactions in germinating maize seeds under salt stress. Journal of Science, Food and Agriculture 92: 839-843.

- Erdal S, 2012b. Alleviation of salt stress in wheat seedlings by mammalian sex hormones. Journal of Science, Food and Agriculture 92: 1411-1416.
- Erdal S, 2012c. Androsterone-induced molecular and physiological changes in maize seedlings in response to chilling stress. Plant Physiology and Biochemistry 57: 1-7.
- Erdal S and Dumlupinar R, 2011a. Mammalian sex hormones stimulate antioxidant system and enhance growth of chickpea plants. Acta Physiologiae Plantarum 33: 1011-1017.
- Erdal S and Dumlupinar R, 2011b. Exogenously treated mammalian sex hormones affect inorganic constituents of plants. Biological Trace Element Research 143: 500-506.
- Fathi S, Kharazmi M and Najafian S, 2019. Effects of salicylic acid foliar application on morphophysiological traits of purslane (*Portulaca olaracea* L.) under salinity stress conditions. Journal of Plant Physiology and Breeding 9(2): 1-9.
- Fazeli Behgo T and Alizadeh Ajirlo S, 2015. Callus induction and shoot regeneration using indole acetic acid and N-isopentenylamino purine combinations and two types of explant in tomato. Journal of Plant Physiology and Breeding 5(2): 41-50.
- Genisel M, Turk H and Erdal S, 2013. Exogenous progesterone application protects chickpea seedlings against chilling-induced oxidative stress. Acta Physiologiae Plantarum 35: 241-251.
- Geuns JMC, 1978. Steroid hormones and plant growth and development. Phytochemistry 17: 1-14.
- Iino M, Nomura N, Tamaki Y, Yamada Y, Yoneyama K and Takeuchi Y, 2007. Progesterone: its occurrence in plants and involvement in plant growth. Phytochemistry 68: 1664-1673.
- Janeczko A, 2012. The presence and activity of progesterone in the plant kingdom. Steroids 77: 169-173.
- Janeczko A and Skoczowski A, 2005. Mammalian sex hormones in plants. Folia Histochemistry and Cytochemistry 43: 71-79.
- Janeczko A and Szybka P, 2001. Induction and proliferation of callus of *Polygonatum verticillatum* L. Conference materials of the 18th Meeting of the Polish Pharmaceutical Society Pharmacy in the 21st century, 19-22 September, Poznan, Poland, p 536 (In Polish).
- Janeczko A, Filek W and Skoczowski A, 2002. Influence of human sex hormones on the growth response of winter wheat immature embryos and callus. Zeszyty Problemowe Postepow Nauk Rolniczych 488: 667-673 (In Polish).
- Janik JR and Adler JH, 1984. Estrogen receptors in Gladiolus ovules. Plant Physiology 75(Suppl): 135.
- Kar M and Mishra D, 1976. Catalase, peroxidase, and polyphenol oxidase activities during rice leaf senescence. Plant Physiology 57: 315-319.
- Khan M, Rozhon W, Unterholzner SJ, Chen T, Eremina M, Wurzinger B, Bachmair A, Teige M, Sieberer T, Isono E and Poppenberger B, 2014. Interplay between phosphorylation and SUMOylation events determines CESTA protein fate in brassinosteroid signaling. Natural Communication 5: 4687 doi:10.1038/ncomms5687.
- Koohi L, Zare N, Asghari Zakaria R and SheikhZadeh-Mosaddegh P, 2014. Effect of plant growth regulators and different explants on tissue culture response and suspension cell cultures of German chamomilla (*Matricaria chamomilla* L.). Journal of Crop Ecophysiology 8: 203-214 (In Persian with English abstract)
- Milanesi L, Monje P and Boland R, 2001. Presence of estrogens and estrogen receptor-like proteins in *Solanum glaucophyllum*. Biochemical and Biophysical Research Communications 289: 1175-1179.
- Murashige T and Skoog F, 1962. A revised method for rapid growth and bioassays with tobacco tissue cultures. Physiologia Plantarum 15: 472-497.
- Petronilho S, Maraschin M, Coimbra MA and Rocha SM, 2012. *In vitro* and *in vivo* studies of natural products: a challenge for their valuation. The case study of chamomile (*Matricaria recutita* L.). Industrial Crops and Products 40: 1-12.
- Pourasadollahi A, Siosemardeh A, Hosseinpanahi F and Sohrabi Y, 2019. Physiological and agromorphological response of potato to drought stress and hormone application. Journal of Plant Physiology and Breeding 9(1): 47-61.
- Shore LS, Kapulnik Y, Ben-Dor B, Fridman Y, Wininger S and Shemesh M, 1992. Effects of estrone and 17β-estradiol on vegetative growth of *Medicago sativa*. Physiologia Plantarum 84: 217-222.
- Simersky R, Novak O, Morris DA, Pouzar V and Strnad M, 2009. Identification and quantification of several mammalian steroid hormones in plants by UPLC-MS/MS. Journal of Plant Growth Regulation 28: 125-136.

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Simons RG and Grinwich DL, 1989. Immunoreactive detection of four mammalian steroids in plants. Canadian Journal of Botany 67: 288-296.

Sudhakar S, Li Y, Katz MS and Elango N, 2001. Translational regulation is a control point in RUNX2/Cbfa1 gene expression. Biochemical and Biophysical Research Communications 289: 616-622.

تأثير ١٧بتا-استراديول بر رشد گياهچه و كالوس در بابونه آلمانی (Matricaria chamomilla L.)

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چکیدہ

واژههای کلیدی: آنزیمهای آنتی اکسیدان؛ القای کالوس؛ کشت درون شیشه ای؛ هورمون استروئیدی